Amendm nts to the Specification:

Please replace the second full paragraph on page 10, lines 12-17, with the following amended paragraph:

Another preferred embodiment of the present aspect of the invention provides that the epitope tag amino acid sequence is selected from the group of YPYDVPDYA (HA) SEQ ID NO:1. EQKLISEEDL (c-*myc*) SEQ ID NO:2, DTYRYI SEQ ID NO:3, TDFYLK SEQ ID NO:4, EEEEYMPME SEQ ID NO:5, KPPTPPPEPET SEQ ID NO:6, HHHHHH SEQ ID NO:7 RYIRS SEQ ID NO:8, or DYKDDDDK SEQ ID NO:9, and that the N-terminus of said protein of interest is fused to a C-terminus of said cell wall binding protein.

Please replace the fifth full paragraph on page 13, lines 23-24, with the following amended paragraph:

Figure 7 shows the sequence of the AGA2-HA-4-4-20-c-myc gene cassette SEQ ID NO:20.

Please replace the second full paragraph on page 17, lines 15-21, with the following amended paragraph:

Figure 18 shows the sequence analysis of wild-type scFv-KJ16 SEQ ID NO:21, mut4 SEQ ID NO:22, and mut7 SEQ ID NO:23. Plasmids from wt scFv-KJ16/yeast and two mutants (mut4 and mut7) were recovered by plasmid rescue and transformed into *E. coli* DH5a competent cells to produce plasmids for sequencing, as described below. Sequence analysis was performed using primers that flank the scFv of the display vector. Mutations are indicated in bold.

Please replace the second full paragraph on page 18, lines 16-18 with the following amended paragraph:

Figure 21 shows the sequence of mutations that lead to the enhanced expression of the cell surface T cell receptor SEQ ID NO:24. These included residues 17 of the Vβ, 43 of the Vα, and 104 of the Vα.

Please replace paragraph 1, page 20, lines 1-2, with the following amended paragraph:

As used herein, the term "HA" refers to the epitope tag sequence YPYDVPDYA SEQ ID NO:1.

Please replace paragraph 2, page 20, lines 3-4, with the following amended paragraph:

As used herein, the term "c-*myc*" refers to the epitope tag sequence EQKLISEEDL SEQ ID NO:2.

Please replace the fourth full paragraph on page 25, lines 23-26, with the following amended paragraph:

A number of polypeptide sequences that can be fused to proteins and bound specifically by antibodies are known and can be utilized as epitope tags. These include, for example, HA SEQ ID NO:1, c-*myc* SEQ ID NO:2, DTYRYI SEQ ID NO:3, TDFYLK SEQ ID NO:4, EEEEYMPME SEQ ID NO:5, KPPTPPPEPET SEQ ID NO:6, HHHHHH SEQ ID NO:7, RYIRS SEQ ID NO:8, and DYKDDDDK SEQ ID NO:9.

Please replace paragraph 4, page 40, lines 7-9, with the following amended paragraph:

- Primers: 5′-ATTAGAATTCCCTACTTCATACATTTTCAA-3′ SEQ ID NO:10 and 5′-ATTACTCGAGCTATTACTGCAGagcgtagtctggaacgtcgtatgggtaAAAAACAT ACTGTGTGTTTATGGG-3′ SEQ ID NO:11.

Please replace the paragraph bridging page 40 and page 41, with the following amended paragraph:

The HA peptide was inserted by cassette mutagenesis. Complementary oligonucleotide strands encoding the Factor Xa recognition sequence and HA epitope were synthesized with cohesive overhangs allowing ligation to the 3′ XhoI site of the AGA2 clone while at the same time destroying this site; a downstream SacI site in pCR-Script annealed and ligated to the CUP1-AGA2 construct in pCR-Script. The insert included a new XhoI site at the 3′end of the HA sequence. CUP1-AGA2-HA was excised as a KpnI/XhoI fragment, purified on a 1% agarose gel, and subcloned into yeast shuttle vector pRS314 (1) already containing the alpha factor terminator sequence, to form surface display vector pCT101. Oligo sequences: 5′-TCGACGATTGAAGGTAGATACCCATACGACGTTCCAGACTACGCTCTGCAGTAATA GATTATCCTCGAGCT-3′ SEQ ID NO:12 and 5′-CGAGGATAATCTATTACTGCAGA GCGTAGTCTGGAACGTCGTATGGGTATCTACCTTCAATCG-3′ SEQ ID NO:13.

Please replace the first full paragraph, page 41, lines 11-17, with the following amended paragraph:

The GAL promoter was excised from vector Ycp1ac22-GAL. 12 bp palindromic linkers with appropriate cohesive overhangs were first cloned into this vector to alter restriction sites at both ends: EcoRI -> KpnI (E/KLINK) and BamHI -> EcoRI (B/ELINK). The resulting KpnI/EcoRI fragment was cloned into pCT101 to form vector pCT201. Oligonucleotide sequences: E/LLINK 5′-AATTGGTACC-3′ SEQ ID NO:14; B/ELINK 5′-GATCGAATTC-3′ SEQ ID NO:15.

Please replace the paragraph 3, page 41, lines 22-24, with the following amended paragraph:

- Primers: 5′-ggttggccaagctagcGACGTCGTTATGACTCAA-3′ SEQ ID NO:16 and 5′-ggccggccaactcgagctattacaagtcttcttcagaaataagcttttgttcTGAGGAGACGGTGAC TGA-3′ SEQ ID NO:17

Please replace the paragraph 4, page 42, lines 5-10, with the following amended paragraph:

The PCR product was cloned into pCR-Script and subcloned into pCT201 as a NheI/XhoI fragment using methods as above, creating vector pCT202. Vector pCT302 was created by inserting a synthetic oligonucleotide (UIUC Biotechnology Center) encoding a $(Gly_4\text{-}Ser)_3$ SEQ ID NO:25 linker in frame between the AGA2 and 4-4-20 open reading frames of pCT202.

Please replace the paragraph 7, page 42, lines 14-15, with the following amended paragraph:

- Primers: 5′-ATTAGAATTCAGCTAAAAAAACCAAAAAAT-3′ SEQ ID NO:18 and 5′-ATTACTCGAGctaTTAACTGAAAATTACATTGC-3′ SEQ ID NO:19

Please replace the second full paragraph on page 52, lines 15-23, with the following amended paragraph:

The AGA2 gene was cloned by PCR from a yeast genomic library and subcloned into an expression vector containing the strong copper-inducible CUP1 promoter which allows 25-fold variation of expression level. Coding sequence for the influenza HA epitope tag (Tyr-Pro-Tyr-Asp-Val-Pro-Asp-Tyr-Ala) SEQ ID NO:1 was fused to the 3′ end of the AGA2 open reading frame, preceded by a Factor Xa site-specific protease

cleavage site (Ile-Glu-Gly-Arg') SEQ ID NO:26. The DNA sequence of this construct is shown in Figure 7. Convenient restriction sites have been included for in-frame fusion of single-chain antibody genes.